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Fluoride Toxicity and Gut Health: Targeting Gut Microbiome and Barrier Function

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ABSTRACT

Fluoride exposure, especially at high doses, can disrupt the gut microbiome by promoting the growth of harmful bacteria while diminishing beneficial microbes, potentially leading to gastrointestinal and systemic health issues. While low fluoride levels may have negligible effects on microbial diversity, excessive fluoride intake can significantly alter gut flora composition, triggering metabolic disruptions and impairing gut barrier function. The review aims to understand how these changes can result in increased intestinal permeability, damage to the epithelial lining, and developing conditions such as leaky gut, where toxins and harmful bacteria pass through the compromised gut lining into the bloodstream. Furthermore, fluoride exposure induces inflammation and oxidative stress, which can aggravate gut dysfunction by affecting the integrity of tight junction proteins essential for maintaining the intestinal barrier. Altered calcium ion homeostasis caused by fluoride also destabilizes these tight junctions and increases intestinal permeability. To address this review, extensive search of peer-reviewed articles was conducted. This review explores the diverse effects of fluoride on gastrointestinal health, focusing on its impact on microbial diversity, intestinal permeability, and inflammatory responses. The evidence suggests that excessive fluoride exposure may lead to significant disruptions in gut health, with long-term consequences such as chronic inflammation, metabolic disturbances, and compromised immune function. A knowledge gap exists in the understanding molecular mechanisms behind these changes. Given the growing concern over fluoride's role in health, particularly in communities with high fluoride exposure, further research is needed to understand its impact on the gut microbiome and overall health outcomes.

Key-words: Fluoride; Gut microbiome; High dose; Intestinal permeability; Leaky gut; Microbial diversity

INTRODUCTION

Organisms are increasingly exposed to various environmental toxic substances, leading to complex biological responses ^[1]. Fluoride (F), a common environmental contaminant, is widespread and poses significant risks to organisms. It is primarily ingested through contaminated water, drugs, toothpaste, and other sources. Drinking water, in particular, is the main route of fluoride exposure, with many countries worldwide reporting elevated fluoride levels in their groundwater ^[2-5]. Research has focused on fluoride's role in dental health, such as cavity prevention, and its potential toxicity at high concentrations, particularly concerning bones, kidneys, liver, and the nervous system ^[6-8]. Several mechanisms have been proposed to explain fluoride-induced diseases, including oxidative stress, signaling pathway alterations, cell cycle disruptions, apoptosis, and epigenetic changes ^[9-10]. A brief outline of fluoride toxicity in humans is shown in Figure 1. While the effects of fluoride on gut health are an emerging area of research, studies in this field remain limited. Alterations in gut microbiota have been linked to various health problems affecting key systems, including the digestive, cardiovascular, nervous, reproductive, and renal systems ^[10-11].

Excessive fluoride intake from drinking water can trigger immune responses, damage the structure of the caecum and rectum, inhibit the proliferation of intestinal epithelial cells, reduce glycoprotein secretion, decrease the number of goblet cells and hypertrophic cells, and alter the diversity and composition of the gut microbiome ^[12,13]. Disruptions in gut microbiota can compromise the integrity of the intestinal barrier, potentially leading to various diseases. Recent studies suggest that disturbances in the gut microbiota and their impact on metabolism and physiological functions play a critical role in disease development ^[14]. Given the known toxicity of fluoride, it is essential to investigate how high levels of fluoride exposure affect the gut microbiota, along with its metabolic and functional changes. The microbiota residing in the gastrointestinal tract plays a crucial role in maintaining overall health, and emerging evidence suggests that fluoride exposure can disrupt the balance of gut bacteria, potentially impairing host health ^[15]. This indicates a complex interaction between

environmental toxicants, gut microbiota, and host health. Excessive fluoride exposure causes significant damage to the rectal structure as well ^[13]. The structural integrity of the intestine is crucial for various biological processes, including digestion, nutrient absorption, and immune defense ^[16]. The intestinal mucosa, which consists of epithelial cells lining the mucosal surface, represents the largest interface between the body and the external environment. As such, the intestinal mucosa is highly susceptible to various biological and environmental factors, which can disrupt its function and even lead to the complete loss of its biological activity ^[17]. Exposure to excessive fluoride notably reduced the thickness of the rectal mucosa and intestinal glands, causing the rectal cavity to either expand or collapse ^[18]. Disruptions were also observed in the arrangement of rectal epithelial cells and the blurring of nuclear membrane boundaries. Interestingly, the thickness of the rectal muscle layer is significantly increased with fluoride due to edema in the intestinal muscle cells and the collapse of the intestinal cavity due to fluoride toxicity ^[19]. The damage to the intestinal mucosa could be linked to oxidative stress and lipid peroxidation ^[20,21]. Fluoride exposure induces cytotoxicity by triggering the generation of reactive oxygen species (ROS) such as superoxide (O_2^-), hydroxyl radicals (OH^\bullet), and hydrogen peroxide (H_2O_2), leading to oxidative stress. This oxidative stress causes lipid peroxidation, DNA damage (including strand breaks and mutations), and disruption of DNA repair mechanisms. As a result, cell membranes become destabilized, leading to cellular dysfunction, apoptosis, and the potential development of cancer. Chronic fluoride exposure has been shown to increase ROS levels, damage cellular structures, and promote inflammation, which intensifies its harmful effects on cells and tissues ^[22-23]. Given the growing concerns over fluoride contamination and its potential impact on gut health, it is essential to elucidate the underlying mechanisms and the broader implications for public health. While much of the existing research on fluoride focuses on its effects on dental health, bones, and neurological development, the impact of fluoride on the gut remains less explored. Thus this review aims to examine the effects of fluoride on the gut and its wider impact on human health.

METHODOLOGY

We conducted an extensive search of research databases such as PubMed, Medline, and Google

Scholar to collect peer-reviewed articles for the preparation of this article.

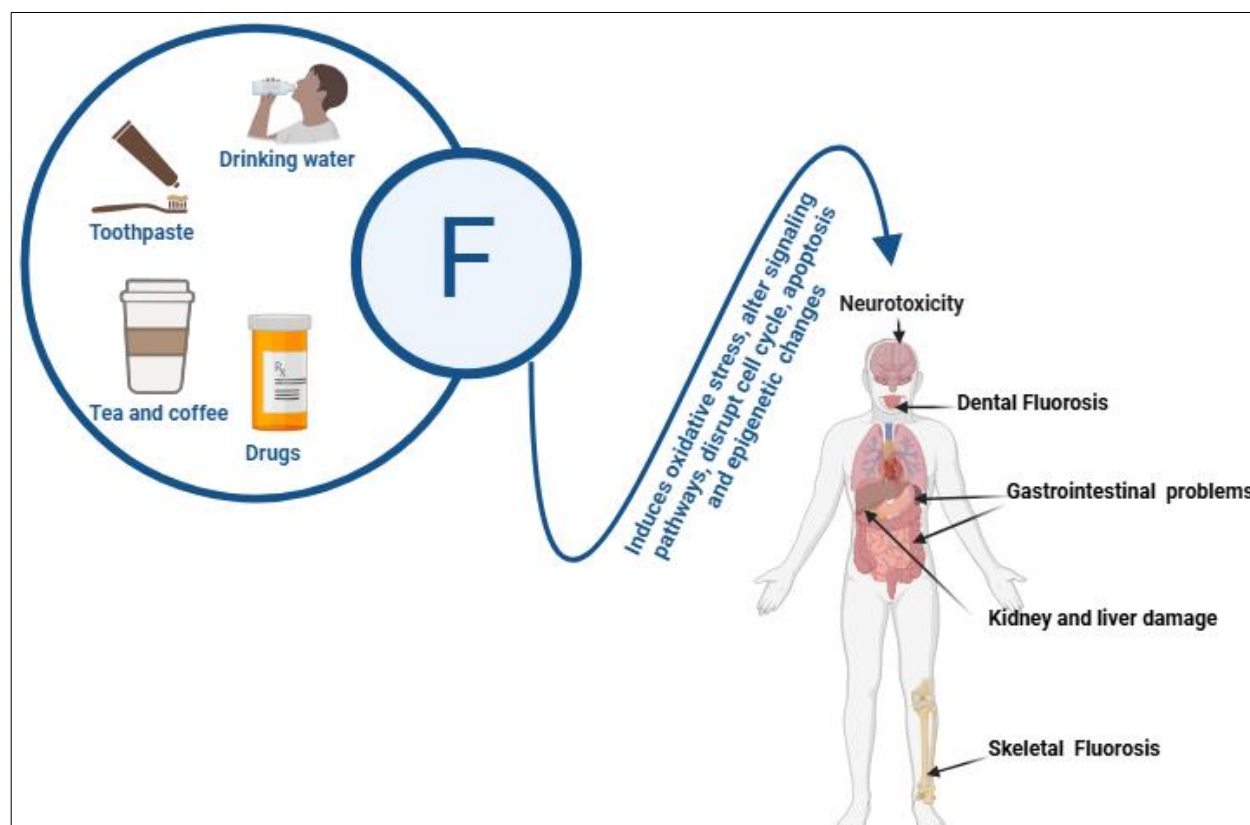


Figure 1. Fluoride toxicity from contaminated water, drugs, toothpaste, and other sources affects bones, kidneys, liver, and the nervous system, with mechanisms involving oxidative stress, signaling pathway alterations, cell cycle disruption, apoptosis, and epigenetic changes.

Fluoride as a Disruptor of the Gut Microbiome

Gut microbiomes and their functions—Our understanding of the microbiome's role in human health has greatly advanced. The gut microbiome, a complex community of bacteria, fungi, and other microorganisms, plays an essential role in digestion, immune function, and overall health [24]. The gastrointestinal (GI) tract hosts around 10^{14} microorganisms, including bacteria, fungi, and viruses, which protect against harmful pathogens, support immune system development, regulate metabolism, and produce vital vitamins [25]. This microbiome begins to form shortly after birth and continues to evolve

throughout life [26]. Maintaining a balanced microbiome in the oral cavity is crucial for preventing dental issues such as caries and periodontal disease [27].

Recent research has suggested that fluoride, particularly when consumed through fluoridated water or toothpaste, could disrupt the balance between the microbiome and its host, leading to potential health issues [28]. Fluoride absorption increases as it moves through the GI tract: about 40% is absorbed in the stomach as hydrofluoric acid (HF), while the remainder is absorbed in the small intestine. Approximately 10% of ingested fluoride is excreted in the feces, indicating exposure of the entire GI tract to some degree of fluoride [29].

Chronic fluoride toxicity places physiological stress on the body [30,31], and evidence suggests that it may also affect the gut microbiome. Studies have shown that ingested fluoride can alter intestinal symptoms and microbial communities in animal models, with high fluoride concentrations reducing beneficial bacteria while promoting the growth of harmful bacteria in vitro fecal fermentation model [32,33]. Additionally, acidogenic oral microbiota, which contributes to tooth decay, has been identified as a major factor in dental fluorosis [34]. Other studies have also linked depleted carbohydrate metabolism pathways in fecal microbiota to an increased risk of dental fluorosis in children [35,36].

Intestinal microflora, a key component of the intestinal mucosal barrier, plays a crucial role in regulating nutrient absorption and preventing the growth of harmful microorganisms. Disruption of the intestinal microflora has been linked to intestinal mucosal barrier dysfunction. It is associated with various diseases, including ulcerative colitis, Crohn's disease, obesity, liver diseases, and immune system disorders [37]. The effects of fluoride on gut microbiota have produced mixed results in research. Some studies have reported that exposure to fluoride concentrations as high as 100 mg/L increases the diversity and richness of the gut microbiome in Kunming and Institute of Cancer Research (ICR) mice [12]. However, other research found no significant changes in the composition or function of the microbiota in BALB/c mice exposed to a 4 ppm fluoride dose [38]. Population studies have shown that children with dental fluorosis, particularly in high-fluoride areas, tend to have slightly lower bacterial diversity and richness than children without fluorosis [39]. Although most studies focus on mice, some evidence suggests that rats have a microbiota more similar to humans, making them valuable models for such research [40]. Most studies have primarily examined the impact of fluoride on microbiota diversity, with fewer exploring the dose-response relationship between fluoride exposure and changes in the microbiota. The networks of metabolic, signaling, and immune-inflammatory axes connect the gut to other organs such as the brain and liver [41].

Recent studies have demonstrated that high fluoride doses, ranging from 45 to 100 mg/L, can significantly alter the gut microbiota in a dose-dependent manner [12]. Fecal samples from healthy individuals aged 20–30 years demonstrated an increased relative abundance

of *Proteobacteria*, a phylum containing several pathogens such as *Escherichia*, *Yersinia*, *Rickettsia*, *Shigella*, *Brucella*, and *Salmonella*, when exposed to high levels of fluoride in the fermentation model. These pathogens are linked to a range of diseases [33]. An overabundance of *Proteobacteria* is increasingly seen as an indicator of dysbiosis, which can lead to disease [42]. Fluoride exposure also significantly impacts gut microbiota at the genus level. *Faecalibacterium*, a key bacterium in the healthy human gut, plays an important role in overall health, with *F. prausnitzii* showing potential as a probiotic [43]. Similarly, *Phascolarctobacterium* has been linked to successful weight loss [44]. However, high fluoride exposure resulted in a marked decrease in the relative abundances of both *Faecalibacterium* and *Phascolarctobacterium*, which may contribute to health risks [33]. A similar decrease in *Faecalibacterium* was observed in mice treated with high fluoride doses [45]. *Firmicutes*, *Bacteroidetes*, and *Proteobacteria* dominant microbial phyla in rodent intestines are decreased fluoride exposure [46]. Within this phylum, *Lactobacillaceae* help lower intestinal pH by producing organic acids and inhibit harmful bacterial growth by altering the permeability of the cytomembrane [47]. The decline in *Lactobacillaceae* following fluoride exposure likely increased the risk of pathogenic infections. Additionally, the relative abundance of *Bacteroidetes* and *Proteobacteria* increased, further exacerbating the imbalance in the intestinal microflora [48]. Furthermore, fluoride treatment led to a significant decrease in the proportions of *Actinobacteria* and *Verrucomicrobia*. *Bifidobacterium*, a key member of *Actinobacteria*, plays a beneficial role in regulating the intestinal microbiome, and its metabolites can inhibit the growth of harmful bacteria such as *Clostridium perfringens* and *Pseudomonas aeruginosa* [48,49]. *Akkermansia*, a member of *Verrucomicrobia*, helps maintain the integrity of the intestinal mucosa by releasing signaling molecules that prevent the passage of lipopolysaccharides [50]. This microbial change could contribute to the damage of the intestinal barrier following fluoride exposure. *Akkermansia*, which interacts with mucin, is decreased with excessive fluoride [51].

Conversely, low fluoride doses have been shown to increase the relative abundance of beneficial bacteria, such as *Lactobacillus* and *Faecalibacterium*. *Lactobacillus* species, widely recognized as beneficial probiotics, play an important role in preventing and

treating various diseases. Many species of *Lactobacillus* are commercially used as probiotics, according to the U.S. Food and Drug Administration ^[52]. Thus, low doses of fluoride may support the growth of beneficial gut bacteria, potentially enhancing host health. In contrast, high fluoride exposure primarily increased the abundance of *Enterobacteriaceae*, a family of intestinal and systemic pathogens, including *Escherichia coli*, *Shigella*, *Klebsiella*, and *Salmonella* ^[33]. These pathogens are associated with chronic inflammation and colorectal cancer. Miao et al. ^[53] found that high sodium fluoride intake altered the cecal microbial community in laying hens by increasing the relative abundance of *Gammaproteobacteria*, *Escherichia-Shigella*, *Streptococcaceae*, and *Enterobacteriaceae*, while decreasing *Lactobacillus*. Moreover, studies utilizing high-throughput 16S rRNA gene sequencing have indicated that fungi play an important role in intestinal health and that excessive fluoride exposure can disrupt the species richness of intestinal fungi, potentially leading to adverse gastrointestinal symptoms by causing imbalances in the fungal microbiota. While high fluoride concentrations have been shown to disrupt microbial communities and potentially contribute to disease risk, low fluoride doses may have a more beneficial effect by promoting the growth of beneficial bacteria. Fluoride exposure not only disturbs the balance of the gut microbiome but also triggers metabolic disruptions. Alterations in the intestinal microbiota and metabolome are key factors in regulating disease susceptibility and multi-organ damage following excessive fluoride exposure. Research has increasingly shown significant changes in the abundance of gut bacteria after excessive fluoride intake, highlighting the role of fluoride in altering the composition of intestinal microbes in animals. Future research is needed to understand better the mechanisms behind these changes and their implications for human health.

Effect of Fluoride on intestinal mucosa

The intestine is a crucial organ in the human body, playing vital roles in food digestion and absorption, maintaining water and electrolyte balance, and regulating immune responses. It absorbs essential nutrients, helps maintain plasma osmotic pressure, and uses specific and non-specific immunity to detect and eliminate harmful pathogens, thereby protecting the body's internal environment. Individuals residing in

areas with high fluoride concentrations often experience gastrointestinal symptoms such as loss of appetite, nausea, bloating, constipation, and intermittent diarrhea. A prospective case-controlled study by Das et al. ^[54] investigated the long-term effects of fluoride ingestion on the gastrointestinal system, revealing significant gastrointestinal disturbances and structural abnormalities. Fluoride-exposed individuals reported symptoms including abdominal pain, vomiting, and nausea. Additionally, petechiae, erosions, and erythema of the mucous membranes were observed in these subjects. Epithelial damage, characterized by a cracked-clay appearance, diminished microvilli, and desquamation, was noted. A report by Spak et al ^[55] highlighted that dental treatment with 0.42%-fluoride gel led to gastric irritation due to accidental swallowing of the gel. This irritation was attributed to the direct toxic effects of fluoride on the gastric mucosa. Petechiae and erosions were observed in the mucosa and surface epithelium was identified as the most affected area of the mucosa. Patients undergoing prolonged fluoride therapy may experience gastric damage unless luminal acidity is reduced with inhibitors or neutralized with antacids. Fluoride acts as a barrier-disrupting agent, promoting ultrafiltration of fluid from the interstitial space into the gastric lumen, which is accompanied by increased acid back-diffusion, a release of glycoproteins, and a reduction in adherent mucus ^[56]. The rapid penetration of fluoride into the mucosa as hydrofluoric acid may aggravate this process, as it induces local vascular stasis, leading to increased intramucosal acidity and, ultimately, significant mucosal damage. The gastrointestinal impact of fluoride is mainly influenced by the concentration of aqueous fluoride in the stomach, rather than the overall fluoride dose from the ingested liquid or solid, emphasizing the importance of fluoride intake concentration in determining health effects.

The fluoride concentration that the gut epithelium is exposed to can be altered by gastric fluids when fluoride is ingested. The population is expected to experience GI symptoms in regions with endemic fluorosis. Therefore, understanding the biological and physiological mechanisms behind fluoride's impact on the GI system is essential. Ingesting high doses of fluoride ^[57-60], such as from fluoride-rich oral care products, contaminated drinking water during fluoridation accidents, or fluoride drugs used for

osteoporosis treatment, can worsen GI symptoms. Fluoride has been shown to stimulate stomach acid secretion, reduce blood flow from the stomach lining, dilate blood vessels, increase redness of the stomach lining, and cause cell death and shedding of the GI tract epithelium. Since fluoride inhibits several key intracellular enzymes, it is not surprising that at high exposures, it causes cell death and desquamation of the GI epithelium. The mechanisms by which fluoride affects secretion are not entirely understood, but they are likely linked to fluoride's ability to activate guanine nucleotide regulatory proteins (G proteins) [61]. Whether fluoride activates these G proteins in the gut epithelium at chronic exposure of fluoridated water at 4.0 mg/L, and has significant effects on gut cell function, remains unclear [62].

Fluoride-Related Inflammation in Gut

The small intestine is one potential site affected by fluoride. Inflammation is known to increase epithelial permeability and inflammatory cytokines such as Interleukin 1 beta (IL-1 β), Tumor necrosis factor alpha (TNF- α), and Interferon gamma (IFN- γ) are associated with enhanced gut permeability. Conversely, IL-10, an anti-inflammatory cytokine, plays a crucial role in maintaining gut homeostasis [63]. Excessive fluoride intake leads to intestinal inflammation by elevating pro-inflammatory cytokines (TNF- α , IL-1 β , and IFN- γ) and reducing the anti-inflammatory cytokine IL-10 [30]. High concentrations of fluoride have been linked to inflammation. In the digestive system, inflammation could play a role in conditions like Irritable Bowel Syndrome (IBS), Crohn's disease, and ulcerative colitis [64]. Chronic excessive fluoride harms the intestine by inhibiting epithelial cell proliferation, increasing intestinal permeability, disrupting intestinal mechanical and immune barrier functions, and promoting inflammation in the gut lining [64]. Furthermore, fluoride-induced oxidative damage can lead to a reduction in the percentage of specific T cell subgroups in the cecal tonsil and decrease the levels of Immunoglobulin A (IgA), Immunoglobulin G (IgG), and Immunoglobulin M (IgM) in this region, which compromises local mucosal immune function [65]. Excess fluoride is absorbed through the stomach and intestinal epithelium, where it can inhibit the proliferation of intestinal epithelial cells and mast cells, stimulate inflammatory cell activity, activate lymphocytes, reduce immunoglobulin concentrations,

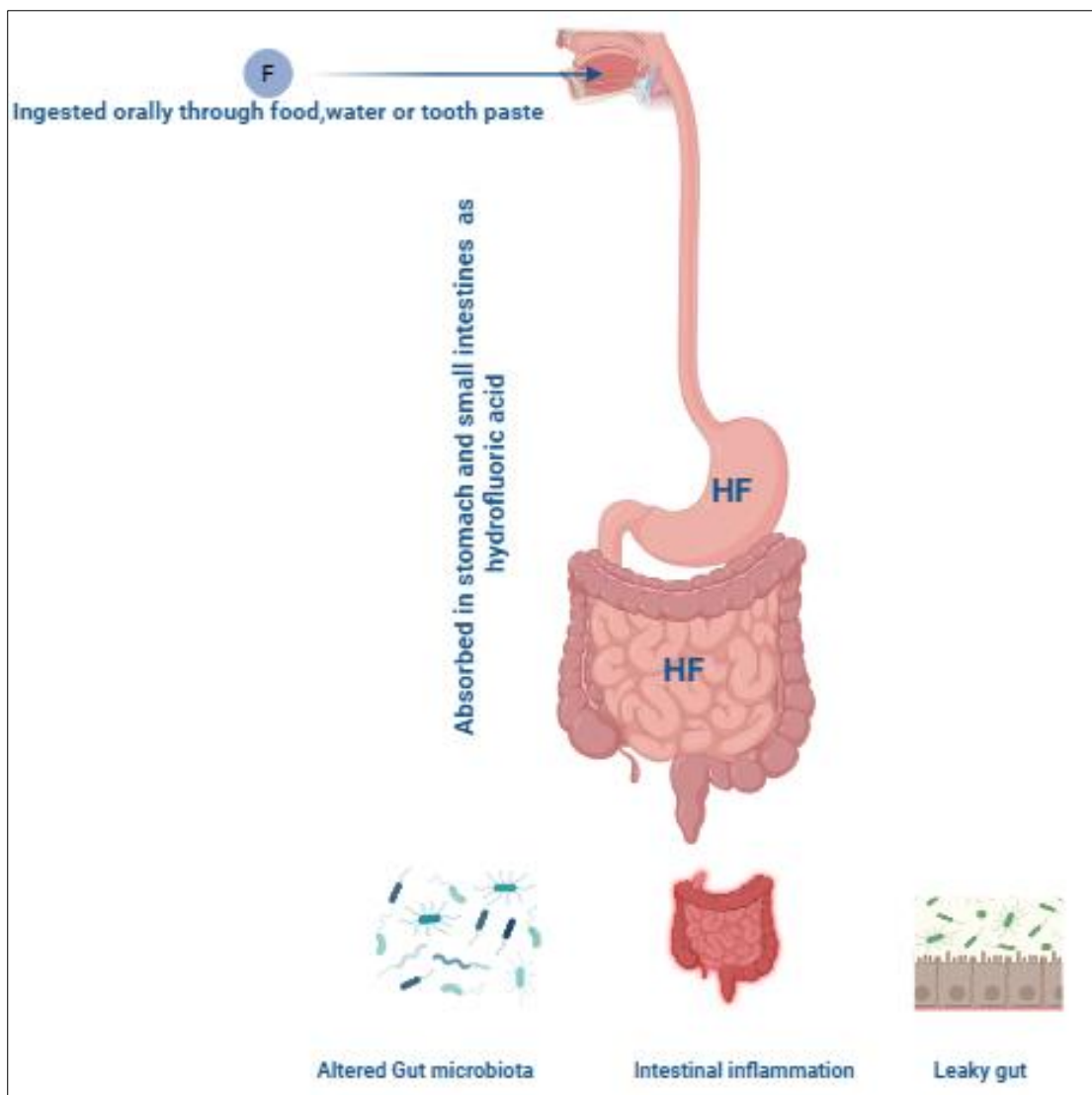
and impair the ability to combat pathogens, ultimately affecting the immune function of the intestinal mucosa [66-69]. The intestinal barrier is a critical defense against microbial pathogens that enter the host via the gut.

Excess fluoride exposure has been shown to trigger pro-inflammatory factor expression, decrease the levels of tight junction-related genes and proteins, activate inflammatory responses, promote cell fever, and hinder intestinal development, leading to inflammation and diarrhea [70]. Increased levels of inflammatory cytokines such as TNF- α and IL-1 β , and elevated Nuclear factor 'kappa-light-chain-enhancer' of activated B-cells (NF- κ B) protein expression in the small intestine are reported in mice having fluorinated drinking water. NF- κ B plays a crucial role in mediating the inflammatory response [71]. White blood cells (WBCs) are recruited by inflammatory cytokines in acute and chronic inflammation, making them a reliable marker of inflammation. The positive association between WBC counts and plasma fluoride concentrations suggests a link between fluoride exposure and heightened inflammation. Recently Den et al [72] reported the positive associations between plasma fluoride and neutrophils and monocytes suggesting that fluoride may influence tissue-specific inflammatory responses. Neutrophils, monocytes, and lymphocytes respond to peripheral inflammation, Lymphocytes are involved in antibody production while neutrophils, are recruited to sites of tissue damage in response to inflammation and are later cleared by monocytes. Fluoride ingestion through drinking water leads to inflammatory lesions in the small intestine resembling those found in Crohn's disease in rodents [19, 73]. Increased neutrophils and monocytes are key components in the chronic inflammation seen in Crohn's disease.

Macrophages are the primary source of reactive oxygen species (ROS) in the human body and play a crucial role in the immune response to various pathogens, as well as in inflammation. When exposed to high fluoride concentrations, macrophages experience an increase in ROS production, leading to oxidative stress, which in turn affects the expression of inflammatory factors [74]. Similarly, fluoride exposure in bovine neutrophils results in the formation of neutrophil extracellular traps (NETs), along with elevated ROS levels and reduced antioxidant enzyme activity [75]. This oxidative stress and NET formation may

contribute to immunotoxicity in neutrophils through activation of the p38/MAPK and ERK pathways. T lymphocytes, including CD4+ T helper (Th) cells and

transcription factors like T-bet and GATA3. These factors are key regulators of Th1/Th2 cell development, differentiation, and memory formation. Disruption of



CD8+ cytotoxic T lymphocytes (CTL), are involved in the immune response by regulating the functions of other immune cells. Excessive fluoride exposure has been shown to reduce the number of lymphocytes, particularly decreasing the percentages of CD4+ and CD8+ T cells in the thymus, which can lead to T cell dysfunction. Fluoride also impacts the expression of

the balance between Th1 and Th2 cells can cause immune dysregulation. Furthermore, excessive fluoride intake has been linked to changes in the number of Th17 cells and alterations in the expression of related cytokines, ultimately affecting the immune system's overall function [76].

Figure 2 - Chronic fluoride toxicity can disrupt the gut microbiota, increase gut inflammation, and lead to a leaky gut.

Fluoride and Gut Barrier Function

The intestinal tract is a protective layer of cells that lines the intestines and prevents harmful substances from entering the bloodstream [77]. The gastrointestinal tract is the earliest and most vulnerable site to toxin exposure. Its health is an indispensable factor for the maintenance of host homeostasis. The physical barrier, the key line of defense, is formed by epithelial cells and the Tight Junctions. TJ acts as paracellular gates that impose size- and charge-dependent restrictions on diffusion, constructed by a network of protein interactions in the apical region of the lateral membrane [78]. Existing research has demonstrated that alterations in the continuity and/or number of tight junction proteins control epithelial permeability. Intestinal permeability, often referred to as "intestinal leakage," can allow the translocation of bacteria and their products, which has been implicated in the development of various health disorders. Tight junction (TJ) proteins, such as ZO-1 and occludin, are essential in maintaining the intestinal barrier by facilitating cell-to-cell adhesion and preventing harmful molecules from crossing the epithelial sheet into the bloodstream [79]. Fluoride induces intestinal inflammation and disrupts mucosal integrity, increasing intestinal permeability [80,81]. The mRNA expression of TJ proteins is significantly reduced in fluoride-exposed rodents contributing to gut inflammation and resulting in increased levels of serum diamine oxidase (DAO) activity and D-lactate. DAO activity is primarily associated with the small intestine, and serum DAO is largely derived from this region in mammals [82]. Additionally, D-lactate, which cannot be produced by mammals but originates from gut bacteria, accumulates in the bloodstream when the intestinal barrier is compromised [83]. Both DAO activity and D-lactate levels are slow to metabolize and serve as valuable markers of mucosal integrity [84]. Long-term fluoride intake directly affects the proliferation of intestinal epithelial cells and destroys the intestinal barrier structure. Intestinal integrity is associated with the construction of tight junctions. Fluoride is absorbed by the stomach and intestines in the form of weakly acidic un-dissociated hydrogen fluoride with the intestine absorbing up to 60% or more. Furthermore, epidemiological investigations showed that among patients with long-term fluoride intake, more than 70% exhibited symptoms such as nausea, vomiting, diarrhea

and abdominal pain. The results of duodenal biopsy from patients with otosclerosis who had consumed 30 mg of sodium fluoride daily showed inflammation of the intestinal mucosa [54]. In addition, some animal studies have suggested that fluoride exposure might alter gut barrier function, potentially leading to "leaky gut," a condition in which the gut lining becomes more permeable.

A leaky gut can allow toxins and undigested food particles to pass into the bloodstream, triggering immune responses and inflammation. Sun et al [85] reported the decreased expression of tight junction protein zonula occludens-1 (ZO-1) in human vascular endothelial cells through the PI3K/AKT signaling pathway through excessive fluoride exposure. Fluoride affects cellular calcium ion homeostasis, with an enhancement in intracellular calcium levels observed in multiple model systems [86]. Fluoride inhibited calcium transporter and channel activity, promoting calcium release from intracellular stores [87]. Xu et al. [88] proved that high fluoride exposure leads to oxidative stress and apoptosis by elevating intracellular calcium concentrations in cultured neuroblastoma cells. Calcium ion is a highly versatile messenger in the modulation of TJs and actin filaments. It is well established that extracellular Ca^{2+} interacts with tight junctions to promote stability. Cytoplasmic calcium is conducive to the assembly of TJs with cytoskeleton. Additionally, calcium ions have been widely reported as mediators of cellular signaling. Ca^{2+} activates MLCK to drive actomyosin reorganization, resulting in the disconnection of tight junctions. Additionally, calcium ion oscillations have been proven to modulate the status of the RhoA/ROCK pathway, and forestalling its overload may rescue barrier hyperpermeability caused by ethanol. The permeability of the intestinal barrier primarily depends on the integrity of TJs between adjacent cells. In contrast to adherens junctions, which initiate cell-cell contacts and control tensile force, TJs act as paracellular gates to confine the trafficking of molecules based on their sizes and charges. TJ disruption mirrored epithelial integrity loss, which triggers multiple diseases that endanger health. NaF administration significantly decreased the expression of the scaffolding protein ZO-1. Consistently, NaF, administered at 100 ppm in drinking water, remarkably reduced the mRNA and/or protein levels of ZO-1 in the ileum of mice [89]. Additionally, 1 mM fluoride impeded TJ formation in rat ameloblast HAT-7 cells [90]. TJ

function is also critically governed by subcellular localization. The rearrangement of TJs has been reported to favor barrier hyperpermeability ^[91]. However, most studies on the correlation between fluoride and TJs have been limited to protein quantification. As confocal microscopy can capture subtle changes in target proteins, NaF noticeably interfered with the continuity of ZO-1 distribution. Ochratoxin A-treated IPEC-J2 monolayers exhibited irregular ZO-1 staining patterns and barrier dysfunction. Another study showed that the bradykinin-mediated blood-tumor barrier (BTB) resulted from the redistribution of ZO-1.

CONCLUSIONS

Fluorine is a highly abundant element that poses toxicity to various organisms, ranging from bacteria to humans. However, the precise mechanisms by which eukaryotes protect themselves from fluoride toxicity are still not well understood. Fluoride disrupts numerous cellular processes, but here, we focus on its impact on the gut.

Excessive fluoride exposure may negatively affect gut health by impairing the intestinal mucosal barrier, disrupting the gut microbiome, and promoting inflammation. Specifically, fluoride exposure decreased beneficial bacteria such as *Lactobacillaceae* and *Bifidobacterium*, while increasing harmful bacteria like *Bacteroidetes* and *Proteobacteria*. This microbial imbalance, coupled with reduced levels of tight junction proteins (ZO-1 and occludin), contributed to increased intestinal permeability. Furthermore, fluoride-induced inflammation was marked by higher levels of pro-inflammatory cytokines and a decrease in the anti-inflammatory cytokine IL-10. Although most health guidelines indicate that typical fluoride levels found in drinking water and dental products are generally safe, maintaining a balanced diet rich in fiber, probiotics, and prebiotics can support a healthy gut microbiome and potentially offset the negative effects of environmental toxins like fluoride. Reducing fluoride exposure should be prioritized in areas with high natural fluoride levels in the water. Individuals concerned about fluoride exposure might consider limiting fluoride toothpaste usage (particularly swallowing large amounts) and avoiding fluoride supplements to lower overall exposure. Future research is needed to understand better the

mechanisms behind these changes and their implications for human health.

FUNDING

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CONFLICT OF INTERESTS

None

REFERENCES

- [1] Sokan-Adeaga AA, Sokan-Adeaga MA, Sokan-Adeaga ED, Oparaji AN, Edris H, Tella EO, Balogun FA, Aledeh M, Amubieya OE. Environmental toxicants and health adversities: A review on interventions of phytochemicals. *J Public Health Res.* 2023 Jun 29;12(2):22799036231181226. doi: 10.1177/22799036231181226.
- [2] Dobaradaran S, Mahvi AH, Dehdashti S. Fluoride content of bottled drinking water available in Iran. *Fluoride* 2008;41(1):93
- [3] Nabipour I, Dobaradaran S. Fluoride concentrations of bottled drinking water available in Bushehr, Iran. *Fluoride* 2013;46(2):63-4.
- [4] Shams M, Dobaradaran S, Mazloomi S, Afsharnia M, Ghasemi M, Bahreinie M. Drinking water in Gonabad, Iran: fluoride levels in bottled, distribution network, point of use desalinator, and decentralized municipal desalination plant water. *Fluoride* 2012;45(2):138
- [5] Mahmoud Shams,a Sina Dobaradaran,b,c Sajad Mazloomi,d Mojtaba Afsharnia,a Mehdi Ghasemi,a Masoud Bahreinie. Drinking water in Gonabad, Iran: Fluoride levels in bottled, distribution network, point of use desalinator, and decentralized municipal desalination plant water, *Fluoride*, Volume 45, 2012.
- [6] Al-Daihan S, Ben Bacha A, El-Ansary A, Bhat RS Prenatal bee pollen treatment improves the neurotoxicity in newborn rats during chronic fluoride exposure in relation to propionic acid-induced rodent models of autism. *Fluoride* 2020;53(1):11-22
- [7] Al-Daihan S, Bhat RS Protective Effect of Bee Pollen Against Sodium Fluoride Induced Hepatonephrotoxicity and Serum Electrolyte Changes in Rats. *Fluoride* 2019; 52(1) 9-17
- [8] Lavanya S, Hema Shree K, Ramani P. Fluoride effect on renal and hepatic functions: A comprehensive decade review of In vitro and In vivo studies. *J Oral Biol Craniofac Res.* 2024 Nov-Dec;14(6):735-745. doi: 10.1016/j.jobcr.2024.10.002. Epub 2024 Oct 15. PMID: 39484005; PMCID: PMC11525447.

- [9] Bhat RS, Soliman DA, Al-Daihan S. Sodium fluoride induces oxidative stress in oral bacteria by altering glutathione (GSH) and glutathione S transferase (GST) activity. *Fluoride* 2021; 54 (1):90-96
- [10] Bhat, RS; Singh, R; Bhat, AM ; Al-Daihan, S Fluoride-Induced Stress in Relation to Brain Health. *Fluoride* 57 (1)2024
- [11] Afzaal M, Saeed F, Shah YA, Hussain M, Rabail R, Socol CT, Hassoun A, Pateiro M, Lorenzo JM, Rusu AV and Aadil RM (2022) Human gut microbiota in health and disease: Unveiling the relationship. *Front. Microbiol.* 13:999001. doi: 10.3389/fmicb.2022.999001
- [12] Mo Z, Wang J, Meng X, Li A, Li Z, Que W, Wang T, Tarnue KF, Ma X, Liu Y, Yan S, Wu L, Zhang R, Pei J, Wang X. The Dose-Response Effect of Fluoride Exposure on the Gut Microbiome and Its Functional Pathways in Rats. *Metabolites*. 2023 Nov 17;13(11):1159. doi: 10.3390/metabo13111159.
- [13] Wang HW, Miao CY, Liu J, Zhang Y, Zhu SQ, Zhou BH. Fluoride-induced rectal barrier damage and microflora disorder in mice. *Environ Sci Pollut Res Int*. 2020 Mar;27(7):7596-7607. doi: 10.1007/s11356-019-07201-8. Epub 2019 Dec 29. PMID: 31885060.
- [14] Hrnecir T. Gut Microbiota Dysbiosis: Triggers, Consequences, Diagnostic and Therapeutic Options. *Microorganisms*. 2022 Mar 7;10(3):578. doi: 10.3390/microorganisms10030578. PMID: 35336153; PMCID: PMC8954387.
- [15] Luo Q, Cui H, Peng X, Fang J, Zuo Z, Deng J, Liu J, Deng Y (2016) Dietary high fluorine alters intestinal microbiota in broiler chickens. *Biol Trace Elem Res* 173(2):483-491. <https://doi.org/10.1007/s12011-016-0672-9>
- [16] Gerritsen J, Smidt H, Rijkers GT, de Vos WM. Intestinal microbiota in human health and disease: the impact of probiotics. *Genes Nutr*. 2011 Aug;6(3):209-40. doi: 10.1007/s12263-011-0229-7.
- [17] Di Tommaso N, Gasbarrini A, Ponziani FR. Intestinal Barrier in Human Health and Disease. *Int J Environ Res Public Health*. 2021 Dec 6;18(23):12836. doi: 10.3390/ijerph182312836. PMID: 34886561; PMCID: PMC8657205.
- [18] Dionizio A, Uyghurturk DA, Melo CGS, Sabino-Arias IT, Araujo TT, Ventura TMS, Perles JVC, Zanon JN, Den Besten P, Buzalaf MAR. Intestinal changes associated with fluoride exposure in rats: Integrative morphological, proteomic and microbiome analyses. *Chemosphere*. 2021 Jun;273:129607. doi: 10.1016/j.chemosphere.2021.129607. Epub 2021 Jan 11. PMID: 33508686; PMCID: PMC8076095.
- [19] Dionizio AS, Melo CGS, Sabino-Arias IT, Ventura TMS, Leite AL, Souza SRG, Santos EX, Heubel AD, Souza JG, Perles J.V.C.M.J.S.r., 2018c. Chronic treatment with fluoride affects the jejunum: insights from proteomics and enteric innervation analysis. 8, 1–12
- [20] Bhat, RS; Aldbass AM; Al-Daihan, S Anti-Biofilm and Antimicrobial Activity of Sodium Fluoride Against Various Pathogenic Microbes. *Fluoride* 57 (1)2024
- [21] Bhat RS , Alonazia MA, Al-Daihana S Fluoride triggers oxidative stress and cell damage in antibiotic-resistant *Klebsiella pneumoniae*. *Fluoride* 57 (11)2024
- [22] He LF, Chen JG. DNA damage, apoptosis and cell cycle changes induced by fluoride in rat oral mucosal cells and hepatocytes. *World J Gastroenterol*. 2006 Feb 21;12(7):1144-8. doi: 10.3748/wjg.v12.i7.1144.
- [23] Chauhan SS, Ojha S, Mahmood A. Modulation of lipid peroxidation and antioxidant defense systems in rat intestine by subchronic fluoride and ethanol administration. *Alcohol*. 2011 Nov;45(7):663-72. doi: 10.1016/j.alcohol.2010.10.008.
- [24] Ma, Z., Zuo, T., Frey, N. et al. A systematic framework for understanding the microbiome in human health and disease: from basic principles to clinical translation. *Sig Transduct Target Ther* 9, 237 (2024). <https://doi.org/10.1038/s41392-024-01946-6>
- [25] Zafar, H., & Saier, M. H. (2021). Gut *Bacteroides* species in health and disease. *Gut Microbes*, 13(1). <https://doi.org/10.1080/19490976.2020.1848158>
- [26] Sedghi L, DiMassa V, Harrington A, Lynch SV, Kapila YL. The oral microbiome: Role of key organisms and complex networks in oral health and disease. *Periodontol* 2000. 2021 Oct;87(1):107-131. doi: 10.1111/prd.12393.
- [27] Rajasekaran, J.J.; Krishnamurthy, H.K.; Bosco, J.; Jayaraman, V.; Krishna, K.; Wang, T.; Bei, K. Oral Microbiome: A Review of Its Impact on Oral and Systemic Health. *Microorganisms* 2024, 12, 1797. <https://doi.org/10.3390/microorganisms12091797>
- [28] Siddiqui R, Badran Z, Boghossian A, Alharbi AM, Alfahemi H, Khan NA. The increasing importance of the oral microbiome in periodontal health and disease. *Future Sci OA*. 2023 Jun 12;9(8):FSO856. doi: 10.2144/fsoa-2023-0062
- [29] Buzalaf MAR, Whitford GM. Fluoride metabolism. *Monogr Oral Sci*. 2011;22:20-36. doi: 10.1159/000325107. Epub 2011 Jun 23. PMID: 21701189.
- [30] Bhat RS, Alghamdi JM, Aldbass AM, Aljebri NA, Alangery AB, Soliman DA, Al-Daihan S Biochemical and FT-IR profiling of *Tritium aestivum* L. seedling in response to sodium fluoride treatment. *Fluoride* 2022; 55 (1): 81-89.
- [31] Bhat RS , Aldbass AM, Alghamdi JM, Alonazia MA, Al-Daihana S *Trigonella foenum-graecum* L seed germination under sodium halide salts exposure. *Fluoride* 2023; 56(2):169-179

- [32] Johnston NR, Strobel SA. Principles of fluoride toxicity and the cellular response: a review. *Arch Toxicol.* 2020 Apr;94(4):1051-1069. doi: 10.1007/s00204-020-02687-5. Epub 2020 Mar 9. PMID: 32152649; PMCID: PMC7230026.
- [33] Chen G, Hu P, Xu Z, Peng C, Wang Y, Wan X, Cai H. The beneficial or detrimental fluoride to gut microbiota depends on its dosages. *Ecotoxicol Environ Saf.* 2021 Feb;209:111732. doi: 10.1016/j.ecoenv.2020.111732. Epub 2020 Dec 26. PMID: 33373928.
- [34] Wang, F., Li, Y., Tang, D. et al. Epidemiological analysis of drinking water-type fluorosis areas and the impact of fluorosis on children's health in the past 40 years in China. *Environ Geochem Health* 45, 9925–9940 (2023). <https://doi.org/10.1007/s10653-023-01772-9>
- [35] Zhou G, Li Q, Hou X, Wu H, Fu X, Wang G, Ma J, Cheng X, Yang Y, Chen R, Li Z, Yu F, Zhu J, Ba Y. Integrated 16S rDNA sequencing and metabolomics to explore the intestinal changes in children and rats with dental fluorosis. *Ecotoxicol Environ Saf.* 2023 Feb;251:114518. doi: 10.1016/j.ecoenv.2023.114518. Epub 2023 Jan 12. PMID: 36640576.
- [36] Spatafora G, Li Y, He X, Cowan A, Tanner ACR. The Evolving Microbiome of Dental Caries. *Microorganisms.* 2024 Jan 7;12(1):121. doi: 10.3390/microorganisms12010121. PMID: 38257948; PMCID: PMC10819217.
- [37] Dmytriv TR, Storey KB, Lushchak VI. Intestinal barrier permeability: the influence of gut microbiota, nutrition, and exercise. *Front Physiol.* 2024 Jul 8;15:1380713. doi: 10.3389/fphys.2024.1380713. PMID: 39040079; PMCID: PMC11260943.
- [38] Yasuda K., Hsu T., Gallini C.A., McLver L.J., Schwager E., Shi A., DuLong C.R., Schwager R.N., Abu-Ali G.S., Franzosa E.A., et al. Fluoride Depletes Acidogenic Taxa in Oral but Not Gut Microbial Communities in Mice. *mSystems.* 2017;2:e00047-17. doi: 10.1128/mSystems.00047-17.
- [39] Wu H. Study on the Characteristics of Gut Microbiota for Children with Dental Fluorosis in Drinking Water-Born Endemic Fluorosis Areas. Zhengzhou University; Zhengzhou, China: 2019.
- [40] Nguyen T.L., Vieira-Silva S., Liston A., Raes J. How informative is the mouse for human gut microbiota research? *Dis. Model Mech.* 2015;8:1–16. doi: 10.1242/dmm.017400.
- [41] Sun, X., Shukla, M., Wang, W. et al. Unlocking gut-liver-brain axis communication metabolites: energy metabolism, immunity and barriers. *npj Biofilms Microbiomes* 10, 136 (2024). <https://doi.org/10.1038/s41522-024-00610-9>
- [42] N.R. Shin, T.W. Whon, J.W. Bae Proteobacteria: microbial signature of dysbiosis in gut microbiota *Trends Biotechnol.*, 33 (9) (2015), pp. 496-503, 10.1016/j.tibtech.2015.06.011
- [43] S. Miquel, R. Martin, O. Rossi, L.G. Bermudez-Humaran, J.M. Chatel, H. Sokol, M. Thomas, J.M. Wells, P. Langella *Faecalibacterium prausnitzii* and human intestinal health *Curr. Opin. Microbiol.*, 16 (3) (2013), pp. 255-261, 10.1016/j.mib.2013.06.003
- [44] D.A.M. Pedrego, M.D. Jensen, C.T. Van Dyke, J.A. Murray, J.A. Woods, J. Chen, P.C. Kashyap, V. Nehra Gut microbial carbohydrate metabolism hinders weight loss in overweight adults undergoing lifestyle intervention with a volumetric diet *Mayo Clinic Proceedings*, 93 (2018), pp. 1104-1110, 10.1016/j.mayocp.2018.02.019
- [45] R. Fu, R. Niu, R. Li, B. Yue, X. Zhang, Q. Cao, J. Wang, Z. Sun Fluoride-induced alteration in the diversity and composition of bacterial microbiota in mice colon *Biol. Trace Elem. Res.* (2019), 10.1007/s12011-019-01942-w
- [46] Wang, Hw., Miao, Cy., Liu, J. et al. Fluoride-induced rectal barrier damage and microflora disorder in mice. *Environ Sci Pollut Res* 27, 7596–7607 (2020). <https://doi.org/10.1007/s11356-019-07201-8>
- [47] Nowak A, Paliwoda A, Błasiak J. Anti-proliferative, pro-apoptotic and anti-oxidative activity of *Lactobacillus* and *Bifidobacterium* strains: A review of mechanisms and therapeutic perspectives. *Crit Rev Food Sci Nutr.* 2019;59(21):3456-3467. doi: 10.1080/10408398.2018.1494539
- [48] Deng F, Zhao B, Yang X et al. The gut microbiota metabolite capsiate promotes Gpx4 expression by activating TRPV1 to inhibit intestinal ischemia reperfusion-induced ferroptosis. *Gut Microbes.* 2021;13(1):1902719 [DOI] [PMC free article] [PubMed] [Google Scholar]
- [49] Huang H, Lin Y, Xin J et al. Fluoride exposure-induced gut microbiota alteration mediates colonic ferroptosis through N6-methyladenosine (m6A) mediated silencing of SLC7A11. *Ecotoxicol Environ Saf.* 2024;283:116816
- [50] Mo C, Lou X, Xue J, Shi Z, Zhao Y, Wang F, Chen G. The influence of Akkermansia muciniphila on intestinal barrier function. *Gut Pathog.* 2024 Aug 3;16(1):41. doi: 10.1186/s13099-024-00635-7. PMID: 39097746; PMCID: PMC11297771.
- [51] Yu, H., Zhang, Y., Zhang, P. et al. Effects of Fluorine on Intestinal Structural Integrity and Microbiota Composition of Common Carp. *Biol Trace Elem Res* 199, 3489–3496 (2021). <https://doi.org/10.1007/s12011-020-02456-6>
- [52] Shah AB, Baiseitova A, Zahoor M, Ahmad I, Ikram M, Bakhsh A, Shah MA, Ali I, Idress M, Ullah R, Nasr FA, Al-Zharani M. Probiotic significance of *Lactobacillus* strains: a comprehensive review on health impacts, research gaps, and future prospects. *Gut Microbes.* 2024 Jan-Dec;16(1):2431643. doi: 10.1080/19490976.2024.2431643.
- [53] Miao L, Zhu M, Li H, Xu Q, Dong X, Zou X. Dietary High Sodium Fluoride Impairs Digestion and Absorption Ability,

- Mucosal Immunity, and Alters Cecum Microbial Community of Laying Hens. *Animals (Basel)*. 2020 Jan 21;10(2):179. doi: 10.3390/ani10020179. PMID: 31973036; PMCID: PMC7070338.
- [54] Das TK, Susheela AK, Gupta IP, Dasarathy S, Tandon RK. Toxic effects of chronic fluoride ingestion on the upper gastrointestinal tract. *J Clin Gastroenterol*. 1994 Apr;18(3):194-9. doi: 10.1097/00004836-199404000-00004. PMID: 8034913.
- [55] Spak CJ, Sjöstedt S, Eleborg L, Veress B, Perbeck L, Ekstrand J. Studies of human gastric mucosa after application of 0.42% fluoride gel. *J Dent Res*. 1990 Feb;69(2):426-9. doi: 10.1177/00220345900690020101.
- [56] Gharzouli K, Amira S, Khenouf S, Gharzouli A. Effects of sodium fluoride on water and acid secretion, soluble mucus and adherent mucus of the rat stomach. *Can J Gastroenterol*. 2000 Jun;14(6):493-8. doi: 10.1155/2000/219623.
- [57] Tangestani, M., Spitz, J., Bahrani, F., Mahvi, A. H., Dobaradaran, S., Ghaedi, H., & Baghmolaei, M. M. (2022). THE NON-CARCINOGENIC RISK OF FLUORIDE VIA CONSUMPTION OF COMMERCIALY AVAILABLE SALT IN GERMANY. *Fluoride*, 55(3), 247-255.
- [58] Darabi, A. H., Tangestani, M., Spitz, J., Bahrani, F., Mahvi, A. H., Heidari, G., & Dobaradaran, S. (2022). FLUORIDE CONTENT OF COMMERCIALY AVAILABLE GREEN TEA IN GERMANY AND ASSESSMENT OF THE NON-CARCINOGENIC RISK. *Fluoride*, 55(2), 145-153.
- [59] Tangestani, M., Mahvi, A. H., Dobaradaran, S., Jamali, M., Saeedi, R., & Spitz, J. (2021). FLUORIDE CONTENT AND HAZARD QUOTIENT FOR BEVERAGES MARKETED IN IRAN. *Fluoride*, 54(2), 178-192.
- [60] Maryam Jamali, Sina Dobaradaran, Amir Hossein Mahv, Alireza Raeisi, Mahbubeh Tangestani, Reza Saeed, Jörg Spitz. Assessing the non-carcinogenic risk due to the intake of fluoride from fruit juice available in the market in Bushehr, *Fluoride*, Volume 53, 2020.
- [61] Strunecka, A.; Strunecky, O. Mechanisms of Fluoride Toxicity: From Enzymes to Underlying Integrative Networks. *Appl. Sci*. 2020, 10, 7100. <https://doi.org/10.3390/app10207100>
- [62] National Academies of Sciences, Engineering, and Medicine. 2006. Fluoride in Drinking Water: A Scientific Review of EPA's Standards. Washington, DC: The National Academies Press. <https://doi.org/10.17226/11571>.
- [63] Di Vincenzo F, Del Gaudio A, Petito V, Lopetuso LR, Scalfaferrì F. Gut microbiota, intestinal permeability, and systemic inflammation: a narrative review. *Intern Emerg Med*. 2024 Mar;19(2):275-293. doi: 10.1007/s11739-023-03374-w. Epub 2023 Jul 28. PMID: 37505311; PMCID: PMC10954893.
- [64] Follin-Arbelet B, Moum B. Fluoride: a risk factor for inflammatory bowel disease? *Scand J Gastroenterol*. 2016 Sep;51(9):1019-24. doi: 10.1080/00365521.2016.1177855. Epub 2016 May 19. Erratum in: *Scand J Gastroenterol*. 2016 Nov;51(11):1. doi: 10.1080/00365521.2016.1229408.
- [65] Liu J, Cui H, Peng X, Fang J, Zuo Z, Deng J, et al. Decreased IgA+ B cells population and IgA, IgG, IgM contents of the cecal tonsil induced by dietary high fluorine in broilers. *Int J Environ Res Public Health*. (2013) 10:1775–85. doi: 10.3390/ijerph10051775 [DOI] [PMC free article] [PubMed] [Google Scholar]
- [66] Gao XY, Jin Y, Zhao J, Zhang YL, Wang HW, Zhou BH. Th17-related cytokines involved in fluoride-induced cecal and rectal barrier damage of ovariectomized rats. *Biol Trace Elem Res*. (2023) 201:4497–507. doi: 10.1007/s12011-022-03519-6
- [67] Liu J, Cui H, Peng X, Fang J, Zuo Z, Wang H, et al. Dietary high fluorine induces apoptosis and alters Bcl-2, Bax, and caspase-3 protein expression in the cecal tonsil lymphocytes of broilers. *Biol Trace Elem Res*. (2013) 152:25–30. doi: 10.1007/s12011-012-9595-2 [DOI] [PubMed] [Google Scholar]
- [68] Chauhan SS, Mahmood A, Ojha S. Ethanol and age enhances fluoride toxicity through oxidative stress and mitochondrial dysfunctions in rat intestine. *Mol Cell Biochem*. (2013) 384:251–62. doi: 10.1007/s11010-013-1804-6
- [69] Cao Q, Li R, Fu R, Zhang X, Yue B, Wang J, et al. Intestinal fungal dysbiosis in mice induced by fluoride. *Chemosphere*. (2020) 245:125617. doi: 10.1016/j.chemosphere.2019.
- [70] Garcia-Hernandez V, Quiros M, Nusrat A. Intestinal epithelial claudins: expression and regulation in homeostasis and inflammation. *Ann N Y Acad Sci*. (2017) 1397:66–79. doi: 10.1111/nyas.13360
- [71] Luo Q, Cui H, Deng H, Kuang P, Liu H, Lu Y, Fang J, Zuo Z, Deng J, Li Y, Wang X, Zhao L. Sodium fluoride induces renal inflammatory responses by activating NF-κB signaling pathway and reducing anti-inflammatory cytokine expression in mice. *Oncotarget*. 2017 Jul 5;8(46):80192-80207. doi: 10.18632/oncotarget.19006. PMID: 29113295; PMCID: PMC5655190.
- [72] Den Besten, P., Wells, C.R. & Abdeweli Uyghurturk, D. Fluoride exposure and blood cell markers of inflammation in children and adolescents in the United States: NHANES, 2013–2016. *Environ Health* 21, 102 (2022). <https://doi.org/10.1186/s12940-022-00911-6>
- [73] Melo CGdS, Perles JVCm, Zanoni JN, Souza SRGD, Santos EX, Leite ADL, Heubel AD, de Souza CO, Souza JGD, Buzalaf MAR. Enteric innervation combined with proteomics for the evaluation of the effects of chronic fluoride exposure on the duodenum of rats. *Sci Rep*. 2017;7(1):1070. doi: 10.1038/s41598-017-01090-y.

- [74] O'Brien KL, Finlay DK. Immunometabolism and natural killer cell responses. *Nat Rev Immunol.* (2019) 19:282–90. doi: 10.1038/s41577-019-0139-2
- [75] Li Y, Du X, Zhao Y, Wang J, Wang J. Fluoride can damage the spleen of mice by perturbing th1/th2 cell balance. *Biol Trace Elem Res.* (2021) 199:1493–500. doi: 10.1007/s12011-020-02264-y
- [76] Zhao Y, Li Y, Wang J, Manthari RK, Wang J. Fluoride induces apoptosis and autophagy through the IL-17 signaling pathway in mice hepatocytes. *Arch Toxicol.* (2018) 92:3277–89. doi: 10.1007/s00204-018-2305-x
- [77] English, J., Connolly, L. & Stewart, L.D. Increased Intestinal Permeability: An Avenue for the Development of Autoimmune Disease?. *Expo Health* 16, 575–605 (2024). <https://doi.org/10.1007/s12403-023-00578-5>
- [78] Lee DB, Jamgotchian N, Allen SG, Abeles MB, Ward HJ. A lipid-protein hybrid model for tight junction. *Am J Physiol Renal Physiol.* 2008 Dec;295(6):F1601-12. doi: 10.1152/ajprenal.00097.2008. Epub 2008 Aug 13.
- [79] Kuo WT, Odenwald MA, Turner JR, Zuo L. Tight junction proteins occludin and ZO-1 as regulators of epithelial proliferation and survival. *Ann N Y Acad Sci.* 2022 Aug;1514(1):21-33. doi: 10.1111/nyas.14798. Epub 2022 May 17. PMID: 35580994; PMCID: PMC9427709.
- [80] Li L, Xin J, Wang H, Wang Y, Peng W, Sun N, Huang H, Zhou Y, Liu X, Lin Y, Fang J, Jing B, Pan K, Zeng Y, Zeng D, Qin X, Bai Y, Ni X. Fluoride disrupts intestinal epithelial tight junction integrity through intracellular calcium-mediated RhoA/ROCK signaling and myosin light chain kinase. *Ecotoxicol Environ Saf.* 2023 Jun 1;257:114940. doi: 10.1016/j.ecoenv.2023.114940. Epub 2023 Apr 24. PMID: 37099960.
- [81] Xin J, Zeng D, Wang H, Sun N, Khalique A, Zhao Y, Wu L, Pan K, Jing B, Ni X. *Lactobacillus johnsonii* BS15 improves intestinal environment against fluoride-induced memory impairment in mice—a study based on the gut-brain axis hypothesis. *PeerJ.* 2020 Oct 7;8:e10125. doi: 10.7717/peerj.10125. PMID: 33083147; PMCID: PMC7547597.
- [82] Luk, Bayless & Baylin (1980). Luk GD, Bayless TM, Baylin SB. Diamine oxidase (histaminase): a circulating marker for rat intestinal mucosal maturation and integrity. *Journal of Clinical Investigation.* 1980;66(1):66–70. doi: 10.1172/JCI109836.
- [83] Sun et al. (2001). Sun XQ, Fu XB, Zhang R, Lu Y, Deng Q, Jiang XG, Sheng ZY. Relationship between plasma D(-)-lactate and intestinal damage after severe injuries in rats. *World Journal of Gastroenterology.* 2001;7(4):555–558. doi: 10.3748/wjg.v7.i4.555.
- [84] Ewaschuk, Naylor & Zello (2005). Ewaschuk JB, Naylor JM, Zello GA. D-lactate in human and ruminant metabolism. *Journal of Nutrition.* 2005;135(7):1619–1625. doi: 10.1093/jn/135.7.1619.
- [85] Sun L; Wang L ; YUAN L . Excessive fluoride exposure decreased the expression of tight junction protein zonula occludens-1, in human vascular endothelial cells Chinese Journal of Endemiology ; (12): 288-292, 2019.
- [86] Aulestia FJ, Groeling J, Bomfim GHS, Costiniti V, Manikandan V, Chaloemtoem A, Concepcion AR, Li Y, Wagner LE 2nd, Idaghdour Y, Yule DI, Lacruz RS. Fluoride exposure alters Ca²⁺ signaling and mitochondrial function in enamel cells. *Sci Signal.* 2020 Feb 18;13(619):eaay0086. doi: 10.1126/scisignal.aay0086.
- [87] Matsuo S, Nakagawa H, Kiyomiya K, Kurebe M. Fluoride-induced ultrastructural changes in exocrine pancreas cells of rats: fluoride disrupts the export of zymogens from the rough endoplasmic reticulum (rER). *Arch Toxicol.* 2000 Feb;73(12):611-7. doi: 10.1007/s002040050015.
- [88] Xu Z, Xu B, Xia T, He W, Gao P, Guo L, Wang Z, Niu Q, Wang A. Relationship between intracellular Ca²⁺ and ROS during fluoride-induced injury in SH-SY5Y cells. *Environ Toxicol.* 2013 Jun;28(6):307-12. doi: 10.1002/tox.20721. Epub 2011 Jul 22. PMID: 21786382.
- [89] Xin J, Wang H, Sun N, Bughio S, Zeng D, Li L, Wang Y, Khalique A, Zeng Y, Pan K, Jing B, Ma H, Bai Y, Ni X. Probiotic alleviate fluoride-induced memory impairment by reconstructing gut microbiota in mice. *Ecotoxicol Environ Saf.* 2021 Jun 1;215:112108. doi: 10.1016/j.ecoenv.2021.112108. Epub 2021 Mar 30. PMID: 33799132.
- [90] Rácz R, Földes A, Bori E, Zsembery Á, Harada H, Steward MC, DenBesten P, Bronckers ALJJ, Gerber G, Varga G. No Change in Bicarbonate Transport but Tight-Junction Formation Is Delayed by Fluoride in a Novel Ameloblast Model. *Front Physiol.* 2017 Dec 6;8:940. doi: 10.3389/fphys.2017.00940. PMID: 29375389; PMCID: PMC5770627.
- [91] Rahner C, Mitic LL, Anderson JM. Heterogeneity in expression and subcellular localization of claudins 2, 3, 4, and 5 in the rat liver, pancreas, and gut. *Gastroenterology.* 2001 Feb;120(2):411-22. doi: 10.1053/gast.2001.21736. PMID: 11159882.