

TRIGONELLA FOENUM-GRAECUM L. SEED GERMINATION UNDER SODIUM HALIDE SALTS EXPOSURE

Ramesa Shafi Bhat,^{a,*} Abeer M Aldbass,^a Jihan Mesfer Alghamdia,^a
Mona Awad Alonazia,^a Sooad Al-Daihan^a
Riyadh, Kingdom of Saudi Arabia

ABSTRACT: Germination of seed to seedling is a physiological process that triggers a cascade of biochemical reactions. The quality of irrigation water can directly affect the early growth stages of a plant. Seeds of *Trigonella foenum-graecum* L. (fenugreek) were exposed to three sodium halides, i.e., NaCl (50 & 100 ppm), NaI (50 & 100 ppm), and NaF (50 & 100 ppm) for two weeks. Seeds grown without the addition of sodium halides were taken as a control for comparison. Seedlings were harvested on the 15th day of sowing and the root and shoot length were recorded. Leaves, shoot, and root was separated, encapsulated in potassium bromide, and then loaded for Fourier transform infrared spectroscopy (FTIR) analysis. A slight increase (16.66%↑) in shoot length was observed with 50 ppm NaCl but 100 ppm NaCl did not show any change in seedling length as compared to the control. 50ppm NaI did not show any change in root length but shoot length was slightly decreased (16.66%↓), and 100ppm NaI showed reduced root (12.5%↓) and shoot length (25%↓) as compared to control. The seeds treated with NaF 100 ppm had the lowest shoot (50%↓) and root length (37.5%↓) among all treated and untreated groups. Unique FT-IR spectral patterns in control and treated seedlings indicate some conformational changes in macromolecules with sodium halide treatment. The overall absorption peaks of treated roots and stems were slightly different from the leaves. The effect of treatment on seedlings was found in the following order; NaCl < NaI < NaF.

Keywords - FT-IR; Sodium fluoride; Sodium chloride; Sodium iodide; *Trigonella foenum-graecum* L

INTRODUCTION

The halogens are universally present in nature and act as micro-nutrients for humans, animals, and plants¹⁻³. Among them, fluorine and chlorine have the highest crustal abundance while iodine stands at the lowest⁴. Fluorine and chlorine are found somewhat in high amounts in crops due to being there in agricultural fields from waste-materials⁵⁻⁶ and iodine levels are increased in the soil through iodinated irrigation water⁷. Total fluoride content of soil ranges from 150–400 mg/kg which is mainly because of phosphorous fertilizers containing 1 – 1.5% fluorine⁸. Also, fluoride naturally occurs in form of metal fluoride salt with sodium in clay and soil. Sodium fluoride (NaF) present in water or soil can alter plant physiology, biochemistry, and structural activities^{9,10}. Likewise, chlorine exists in combination with some inorganic substances like sodium. Sodium chloride (NaCl) mainly contributes to salinity stress in agricultural land which is composed of a range of dissolved salts¹¹⁻¹³. Chlorine is able to accumulate in leaf tissue, resulting in leaves with a scorched or burned appearance. Chlorine is present in a small amount and acts as a micronutrient and participates in several physiological processes in plants¹⁴. Iodine is considered a micro-nutrient not only for humans but also for plants. Sodium iodide (NaI) is added to table salt to treat iodine deficiency. The biofortification of crops with iodine is growing these days in order to increase iodine content in plants to

^aBiochemistry Department, Science College of King Saud University, Riyadh, Saudi Arabia. For correspondence: Ramesa Shafi Bhat, Biochemistry Department, Science College of King Saud University, PO Box 22452, Zipcode 11495, Riyadh, Saudi Arabia. E-mail: rbhat@ksu.edu.sa

fulfill the daily intake requirements for humans¹⁵. Iodine is not an essential nutrient for plants and its high levels in irrigation waters can be toxic¹⁶.

The seed germination phase is considered a vital part of the plant's life cycle as the yield of the crops depends fully on the germination process. Elements that are present in water or soils can directly affect the germination process. Recently, Fourier-transform infrared spectroscopy (FTIR) has been used to observe the biochemical changes in plants' response to abiotic stresses in all stages of plant development¹⁷. An acquisition time of FTIR is very short and suitable to examine the initial response of seedlings to stress¹⁸. Fenugreek (*Trigonella foenum-graecum* L.) is known for its medicinal value all over the world. Fenugreek is a fast-growing plant and needs only 5–10 days for germination¹⁹. In the present study, we explored the effect of different sodium halides on seedling growth and biochemical changes during the germination process in fenugreek (*Trigonella foenum-graecum* L) seeds.

MATERIAL AND METHODS

Plant material: Seeds of fenugreek were purchased from the local market in Riyadh, Saudi Arabia, and taxonomic identification was confirmed by Dr. Mona. S. Alwhibi in Botany and Microbiology Department, College of Science, King Saud University. An equal number of seeds were placed in different Petri dishes containing sterile cotton, dispersed in 10 mL of aqueous solutions of different halogen elements. Seeds were germinated for 15 days with different concentrations of aqueous solutions of NaCl, NaI, and NaF as mentioned below

Treatment: The experimental design with the different treatments was arranged below

1. *Control:* Fenugreek seeds germinated in distilled water;
2. *NaCl treated:* Fenugreek seeds germinated in 50 & 100 ppm NaCl aqueous solution
3. *NaI treated:* Fenugreek seeds germinated in 50 & 100 ppm NaI aqueous solution.
4. *NaF treated:* Fenugreek seeds germinated in 50 & 100 ppm NaF aqueous solution.

All Petri dishes were placed in a growth chamber for two weeks. 1 mL of the respective solutions was added to each Petri dish every day in order to maintain adequate moisture.

Growth analysis:

After two weeks of treatment, seedlings were harvested carefully. The root and shoot length was recorded with the help of a ruler. After harvesting, tiny leaves, shoots, and roots were collected separately for Fourier-transform infrared spectroscopy.

Fourier-transform infrared spectroscopy (FTIR): Freshly collected leaves, shoots, and roots were homogenized with distilled water and left to dry. The dried samples were encapsulated in potassium bromide and then loaded onto an FTIR spectroscope and results were recorded on Thermo Scientific-Nicolet -6700 FTIR spectrometer at a scan range of 400–4000 cm^{-1} with a resolution of 4 cm^{-1} .

RESULTS

Effect of different halogen elements on the growth of the seedling

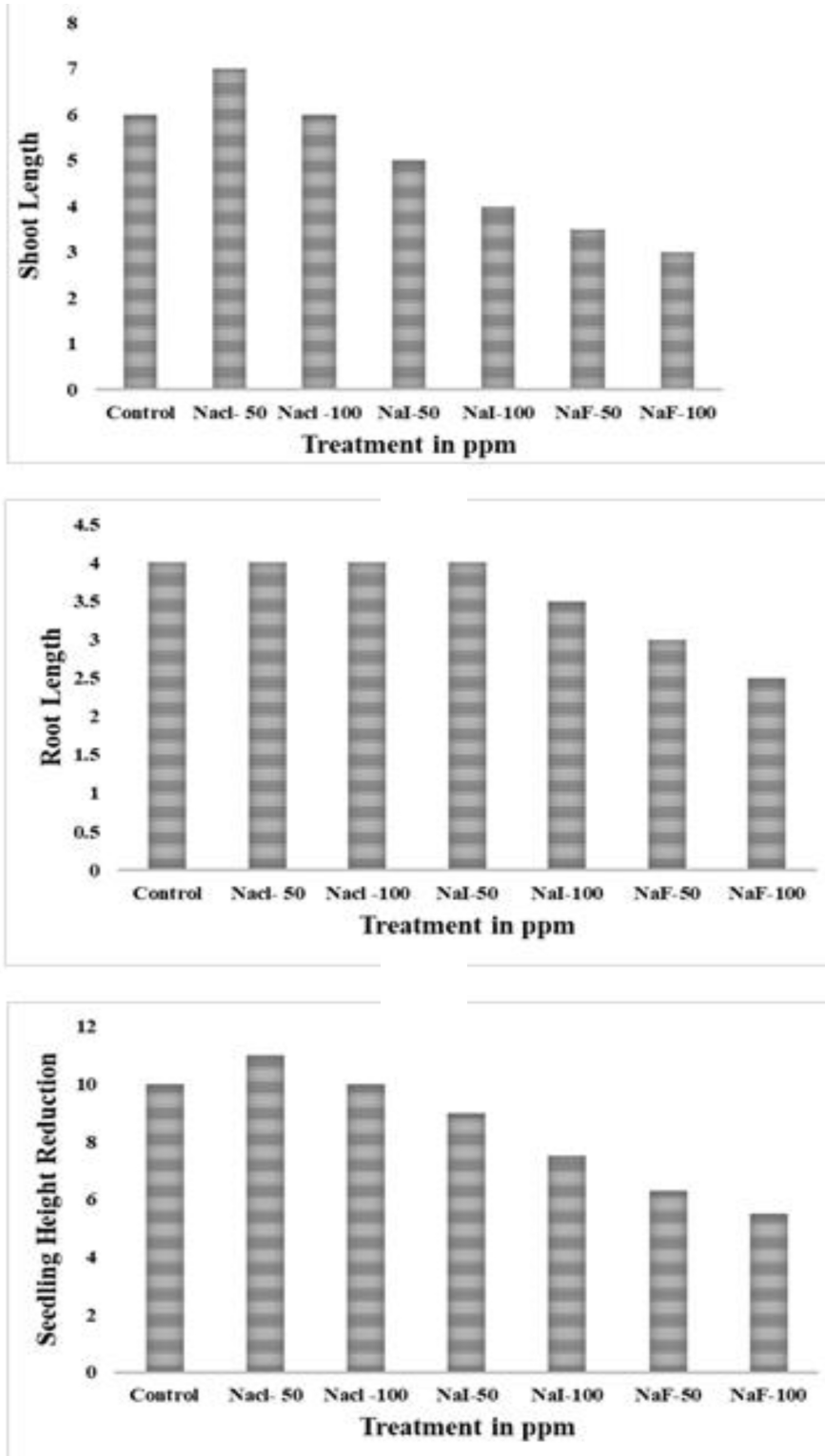
The Table and Figure 1 show the effect of different concentrations of sodium halides on seedling height. We found a slight increase in shoot length with no change in root length in seeds treated with 50 ppm of NaCl while seeds treated with 100 ppm NaCl did not show any change in seedling height when compared with the control. 50 ppm NaI did not show any change in root length but shoot length was slightly decreased, and 100 ppm NaI showed reduced root and shoot length as compared to the control. The seeds treated with NaF (50 & 100 ppm) had the lowest shoot and root length among all treated and untreated groups. As a result, the effect of halogen elements on seedling growth was found in the following order; NaCl < NaI < NaF (Figure 1).

Table. Percentage change of shoot length; root length, and seedling height reduction (SHR) in all groups compared to control

Group	Percent change		
	Shoot length	Root length	SHR
Control	100.00	100.00	100.00
NaCl-50	116.66	100.00	110.00
NaCl-100	100.00	100.00	100.00
NaI-50	83.84	100.00	90.00
NaI-100	75.00	87.50	75.00
NaF-50	66.66	75.00	63.00
NaF-100	50.00	62.50	55.00

Effect of different halogen elements on FTIR analysis

FTIR analysis of roots, shoots, and leaves samples collected from seeding treated with different concentrations of sodium halides, are shown in Figure 2, Figure 3, and Figure 4, respectively. The recorded spectral result shows some unique spectral patterns in control and treated seedling at certain wavenumbers which indicates some conformational changes in macromolecules such as lipids (3000–2800 cm^{-1}), proteins and lignin (1700–1500 cm^{-1}), lipids, and pectin (1790–1720 cm^{-1}), cellulose and hemicellulose (1300–1180 cm^{-1}) and other carbohydrates (fingerprint region 1200–900 cm^{-1}). In the root sample, the differences in band intensities between control and halogen treated plants were prominent at wavenumbers 3980, 3059, 3025, 2921, and 2852 cm^{-1} indicating conformational changes in lipids and 1635, 1601, 1600, 1492, 1449, 1152, 1069, 1027, and 905 cm^{-1} denoted cell wall region (Figure 2). In shoot samples, the same pattern of band shifts was found with few additional bands at 2366 and 2340 cm^{-1} mentioning more changes in lipids. We observed changes in band intensities in leaf tissue at 3389, 3060, 2922, 2853, and 2368 cm^{-1} indicating lipids 1944, 1636, 1605, 1493, 1449, and 1428 cm^{-1} indicating proteins, 1069, 906, 839, 754, 696, 536 cm^{-1} indicating carbohydrates (Figure 3). The average absorbance of lipids, protein, carbohydrates, and cell wall components of sodium halide-treated root, stem, and leaf is shown in Figure 5.



Figures 1A–1C. Effects of different treatment on 1A: shoot length; 1B: Root length; and 1C seedling height reduction.

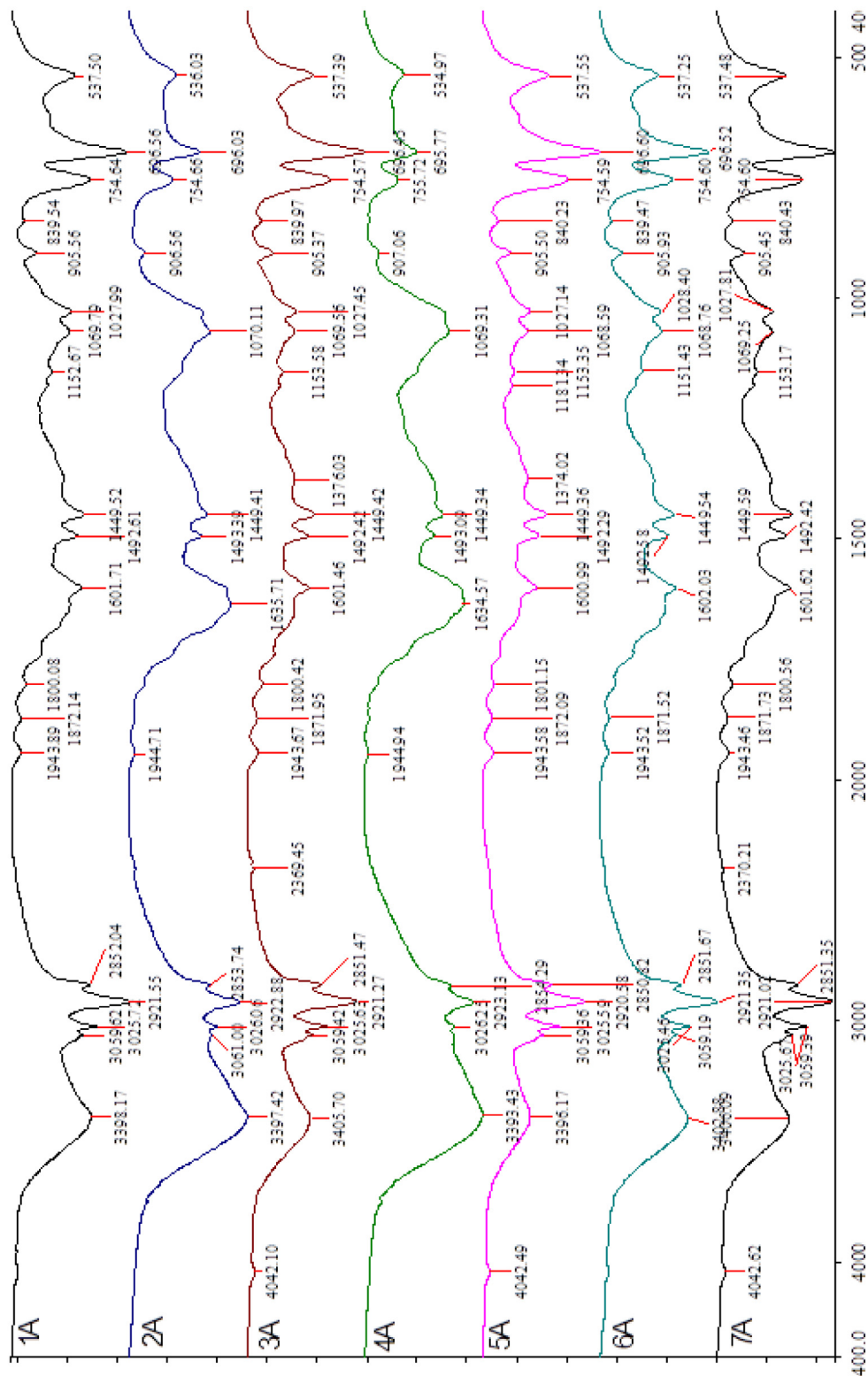


Figure 2. IF-IR spectra of roots of seedlings treated with different halogen elements with concentration in ppm. 1A: control; 2A: NaCl 50; 3A: NaCl 100; 4A: NaI 50; 5A: NaI 100; 6A: NaF 50; and 7A: NaF100.

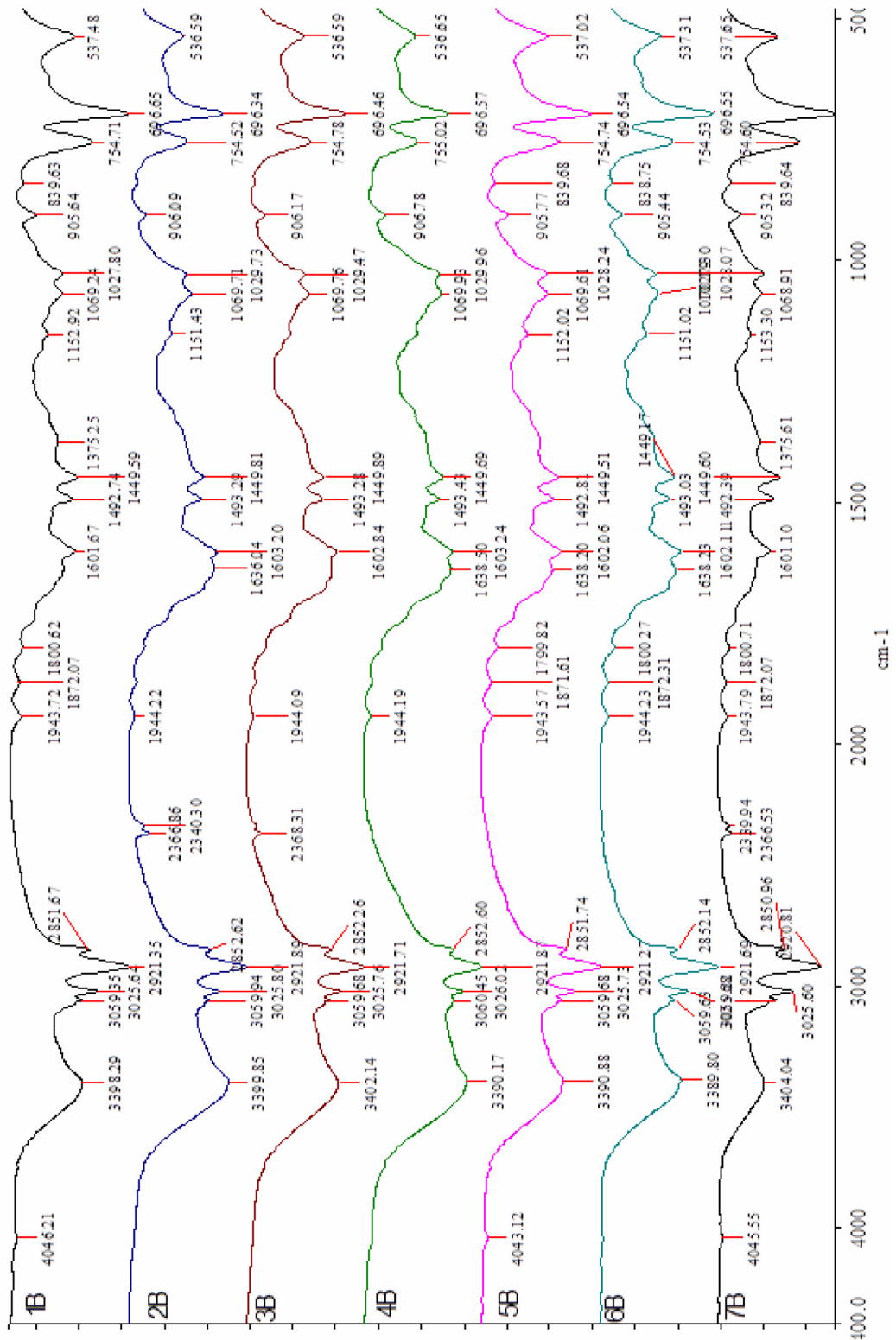


Figure 3. IF-IR spectra of stem of seedlings treated with different halogen elements with concentration in ppm. 1B: control; 2B: NaCl 50; 3B: NaCl 100; 4B: NaI 50; 5B: NaI 100; 6B: NaF 50; and 7B: NaF 100.

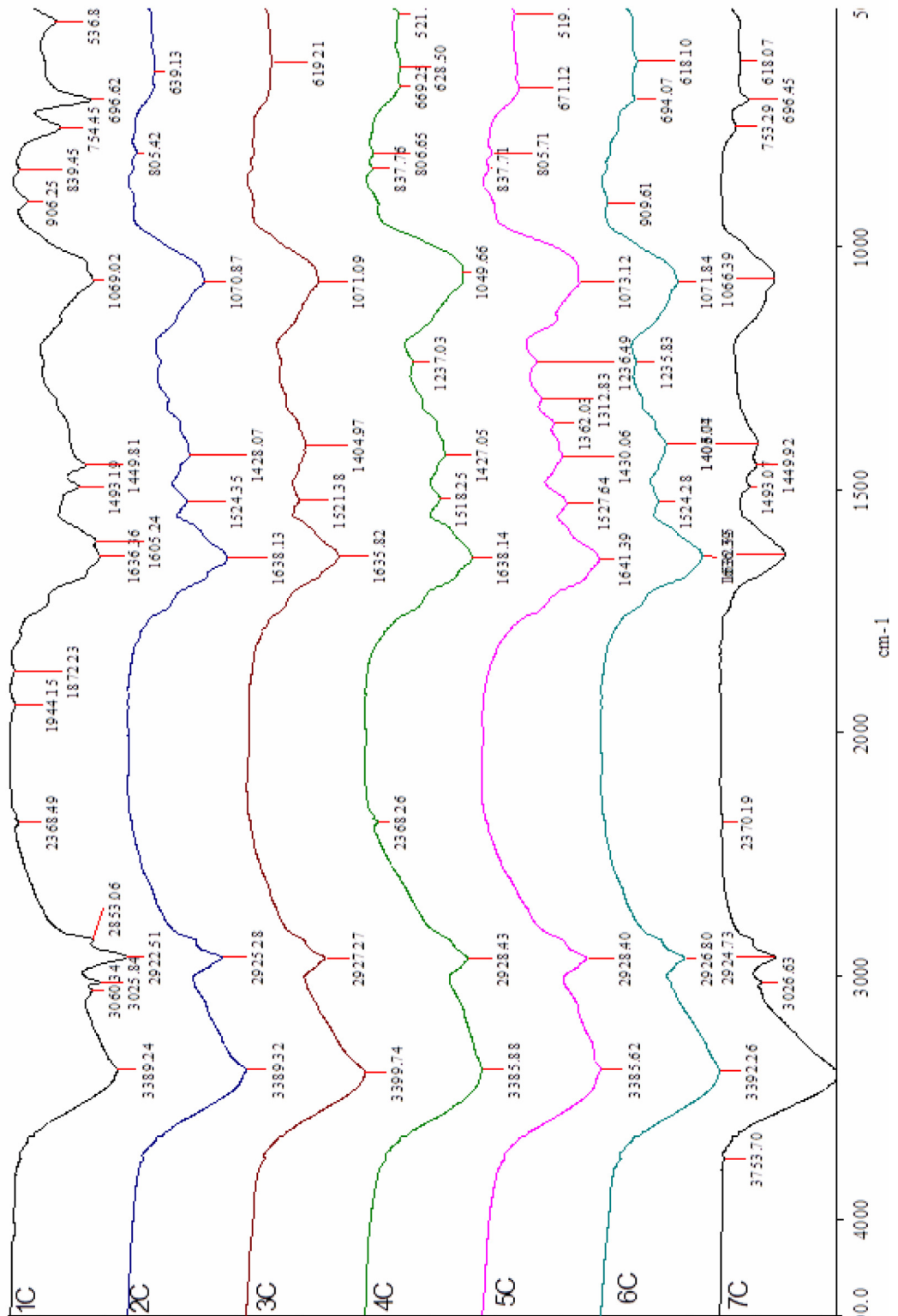


Figure 4. IF-IR spectra of leaves of seedlings treated with different halogen elements with concentration in ppm. 1C: control; 2C: NaCl 50; 3C: NaCl 100; 4C: NaI 50; 5C: NaI 100; 6C: NaF 50; and 7C: NaF100.

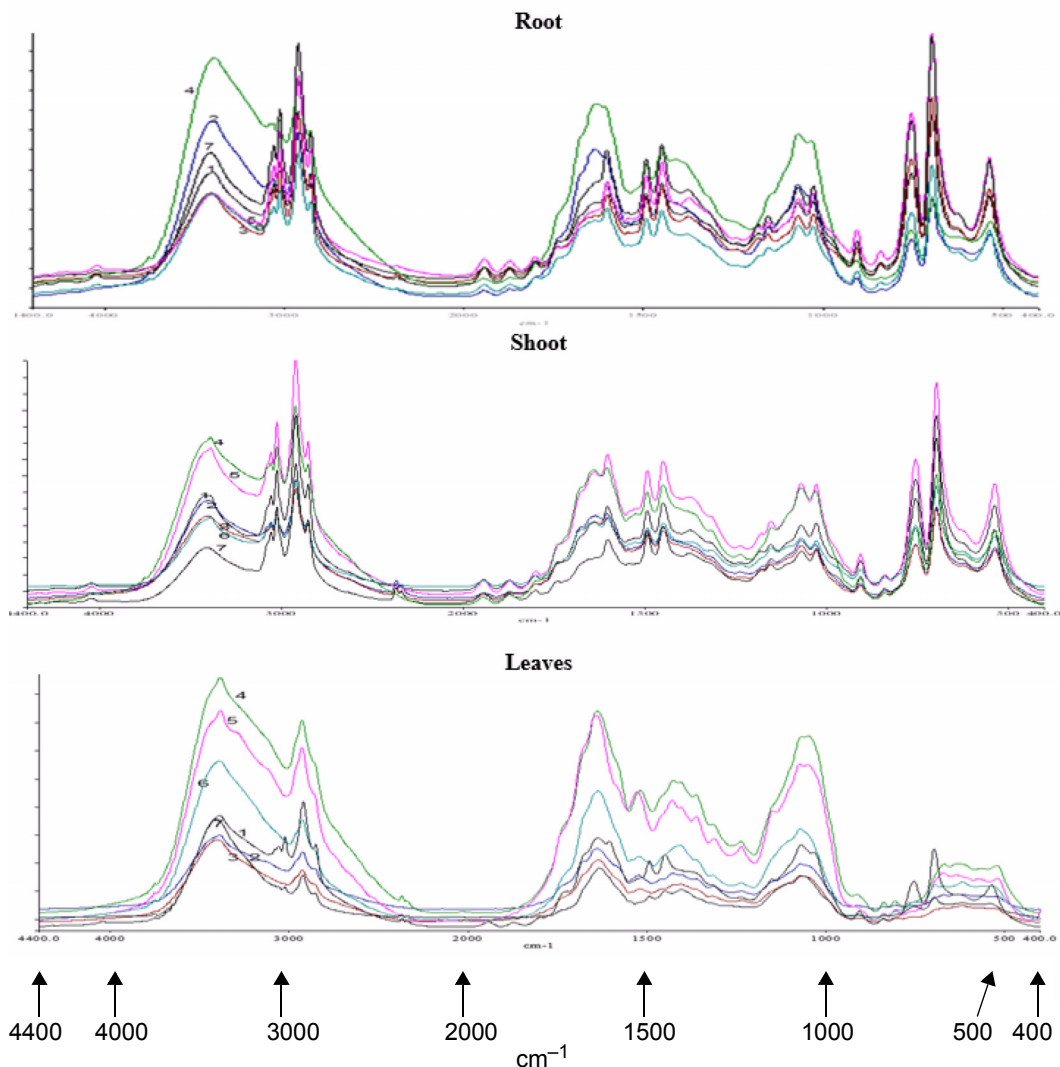


Figure 5. Average lipid (3000–2000 cm^{-1}) absorbance; average proteins (1800–1500 cm^{-1}) absorbance; average carbohydrates (1500–1200 cm^{-1}) and average cell wall components (1000–600 cm^{-1}) of seedlings parts, (root, shoot, and leaves), treated with different halogen elements with concentration in ppm. 1: control; 2: NaCl 50; 3: NaCl 100; 4: NaI 50; 5: NaI 100; 6: NaF 50; and 7: NaF 100.

DISCUSSION

Water is very essential for seed germination. The quality of water used for irrigation can directly affect seedling growth. Fenugreek seed germinated fine with NaCl solutions as compared to NaI and NaF. 100ppm NaCl did not show any change in the root or shoot length but, surprisingly, 50ppm NaCl stimulated the growth of seedlings (Figure 1). NaCl is known to be toxic for plants but not at lower concentrations. NaCl can provide a better ion balance, without causing any change to

cellular metabolism and the root-to-shoot distribution of other essential nutrients²⁰. Improvement in growth at the low level of NaCl in plants has also been reported earlier²¹⁻²³. The same concentration (50 and 100ppm) of NaI and NaF reduced both root and shoot length (Figure 1). Some earlier reports concluded that toxic levels of fluoride and iodine can cause reduce root and shoots length due to imbalanced nutrient uptake by seedling²³⁻²⁵.

FTIR analysis is involved in the identification and quantification of the plant substances like cell wall components, proteins, and lipids in order to understand the depth of biotic and abiotic stresses in plants^{26,27}. Different trends of peaks positions from the spectra in different regions of lipids, proteins, carbohydrates, and cell walls were slightly different in control plants as compared to treated seedlings (Figures 2–5). Biochemical changes in samples can be achieved by analyzing the peak width, position, and intensity of absorption²⁸⁻³⁰. The band height changes indicate some modifications of chemical groups³¹ in lipids, carbohydrates, proteins, and cell wall regions. The absorption peaks of treated roots and stems were slightly different from those of leaves. Lipids, carbohydrate, and protein regions of treated leaves showed different peaks when compared to treated roots and stems. The lipids and cell wall region of roots and shoots showed the almost same pattern. The leaf is very sensitive to water stress whereas the root is more resistant³¹. Quality water is vital for the growth and development of a plant and can bring physical or chemical changes in which many biological molecules such as nucleic acids, proteins, carbohydrates, and lipids are involved³². Higher or lower absorption levels for most wave numbers in NaI-treated and NaF-treated seedlings than for the control indicate an accumulation or reduction of various biochemical components in order to cope with stress³³⁻³⁴.

CONCLUSIONS

Fenugreek seedlings were tolerant to NaCl salinity up to 100 ppm but the same amount of NaI and NaF caused reduced growth and biochemical changes. Changes in leaves were slightly different than root and shoot. FTIR spectroscopy can be used to detect conformational changes of biological molecular components in young seedlings as it only requires small sample sizes.

ACKNOWLEDGMENTS

This research project was supported by Researchers Supporting Project (RSP) number 2023/183, King Saud University, Riyadh, Saudi Arabia.

REFERENCES

- [1] Smolin LA, Grosvenor MB. The trace elements: fluoride. In Nutrition: Science and applications. 2nd ed. New Jersey: John Wiley and Sons; 2010. pp. 515-8.
- [2] Raven JA. Chloride: essential micronutrient and multifunctional beneficial ion. J Exp Bot 2017;68(3):359-67.
- [3] Kiferle C, Martinelli M, Salzano AM, Gonzali S, Beltrami S, Salvadori PA, Hora K, Holwerda HT, Scaloni A, Perata P. Evidences for a nutritional role of iodine in plants. Front Plant Sci 2021;12:616868.
- [4] Fuge R. Sources of halogens in the environment, influences on human and animal health. Environ Geochem Health 1988;10(2):51-61.

- 178 Research report *Trigonella foenum-graecum* L. seed germination under 178
Fluoride 56(2):169-179 sodium halide salts exposure
April-June 2023 Bhat, Albass, Alghamdi, Alonazia, Al-Daihan
- [5] Zhang J, Yu L, Yang H, Ye B. Migration and transformation of fluoride through fluoride-containing water for the irrigation of a soil-plant system. *Hum Ecol Risk Assess: An International Journal* 2019;25(4):1048-58.
- [6] Zaman M, Shahid SA, Heng L. Irrigation water quality. In: *Guideline for salinity assessment, mitigation and adaptation using nuclear and related techniques*. Cham: Springer; 2018.
- [7] Cao XY, Jiang XM, Kareem A, Dou ZH, Abdul Rakeman M, Zhang ML, Ma T, O'Donnell K, DeLong N, DeLong GR. Iodination of irrigation water as a method of supplying iodine to a severely iodine-deficient population in Xinjiang, China. *Lancet* 1994;344(8915):107-10.
- [8] Bombik E, Bombik A, Górski K, Saba L, Bombik T, Rymuza K. The effect of environmental contamination by fluoride compounds on selected horse tissues. *Polish J of Environ Stud* 2011;20(1):37-43.
- [9] Pelc J, Smolik B, Krupa-Matkiewicz M. Effect of sodium fluoride on some morphological and physiological parameters of 10-day-old seedlings of various plant species. *Folia Pomer Univ Technol Stetin Agric Aliment Pisc Zootech* 2017;338:151-8.
- [10] Bhat RS, AlGhamdi JM, Albass AM, Aljebri NA, Alangery AB, Soliman DA, Al-Daihan S. Biochemical and FT-IR profiling of *Triticum aestivum* L seedlings in response to sodium fluoride treatment. *Fluoride* 2022;55(1):81-9.
- [11] Wu GQ, Jiao Q, Shui QZ. Effect of salinity on seed germination, seedling growth and inorganic and organic solutes accumulation in sunflower (*Helianthus annuus* L.). *Plant Soil Environ* 2015; 61:220-6.
- [12] Munns R, Tester M. Mechanisms of salinity tolerance. *Annu Rev Plant Biol* 2008; 59:651-81.
- [13] Geilfus CM. Chloride: from nutrient to toxicant. *Plant Cell Physiol* 2018;59(5):877-86.
- [14] Chen WR, He ZL, Yang XE, Mishra S, Stoffella PJ. Chlorine nutrition of higher plants: progress and perspectives. *J Plant Nutr* 2010;33(7):943-52.
- [15] Dávila-Rangel IE, et al. Iodine biofortification of crops. In: Jaiwal P, Chhillar A, Chaudhary D, Jaiwal R, editors. *Nutritional quality improvement in plants. Concepts and strategies in plant sciences*. Cham: Springer; 2019.
- [16] Kato S, Wachi T, Yoshihira K, Nakagawa T, Ishikawa A, Takagi D, Tezuka A, Yoshida H, Yoshida S, Sekimoto H, Takahashi M. Rice (*Oryza sativa* L.) roots have iodate reduction activity in response to iodine. *Front Plant Sci* 2013;4:227.
- [17] Yang J, Yen HE. Early salt stress effects on the changes in chemical composition in leaves of ice plant and *Arabidopsis*. A Fourier transform infrared spectroscopy study. *Plant Physiol* 2002;130(2):1032-42.
- [18] Strakov P, Larmola T, Andrés J, Ilola N, Launiainen P, Edwards K, Minkinen K, Laiho R. Quantification of plant root species composition in peatlands using FTIR spectroscopy. *Front Plant Sci* 2020;11:597.
- [19] Ahmad A, Alghamdi SS, Mahmood K, Afzal M. Fenugreek a multipurpose crop: Potentialities and improvements. *Saudi J Biol Sci* 2016;23(2):300-10.
- [20] Hongqiao L, Suyama A, Mitani-Ueno N, Hell R, Maruyama-Nakashita A. A low level of NaCl stimulates plant growth by improving carbon and sulfur assimilation in *Arabidopsis thaliana*. *Plants* 2021;10(10):2138.
- [21] Ebrahimi R, Bhatla SC. Effect of sodium chloride levels on growth, water status, uptake, transport, and accumulation pattern of sodium and chloride ions in young sunflower plants. *Commun Soil Sci Plant Anal* 2011;42(7):815-31.

- 179 Research report *Trigonella foenum-graecum* L. seed germination under 179
Fluoride 56(2):169-179 sodium halide salts exposure
April-June 2023 Bhat, Albass, Alghamdia, Alonazia, Al-Daihan
- [22] Cao Y, Liang L, Cheng B, Dong Y, Wei J, Tian X, Peng Y, Li Z. Pretreatment with NaCl promotes the seed germination of white clover by affecting endogenous phytohormones, metabolic regulation, and dehydrin-encoded genes expression under water stress. *Int J Mol Sci* 2018;19(11):3570.
- [23] Sogoni A, Jimoh MO, Kambizi L, Laubscher CP. The impact of salt stress on plant growth, mineral composition, and antioxidant activity in *Tetragonia decumbens* Mill.: An underutilized edible halophyte in South Africa. *Horticulturae* 2021;7(6):140.
- [24] Sabal D, Khan TI, Saxena R. Effect of sodium fluoride on cluster bean (*Cyamopsis tetragonoloba*) seed germination and seedling growth. *Fluoride* 2006;39(3):228-30.
- [25] Zhu YG, Huang YZ, Hu Y, Liu YX. Iodine uptake by spinach (*Spinacia oleracea* L.) plants grown in solution culture: effects of iodine species and solution concentrations. *Environ Int* 2003;29(1):33-7.
- [26] Wang J, Zhu J, Huang RZ, Yang YS. Investigation of cell wall composition related to stem lodging resistance in wheat (*Triticum aestivum* L.) by FTIR spectroscopy. *Plant Signaling and Behavior* 2012; 7: 856-63.
- [27] Conrad AO, Bonello P. Application of infrared and raman spectroscopy for the identification of disease resistant trees. *Front Plant Sci* 2016; 6
- [28] Gorzsas A, Stenlund H, Persson P, Trygg J, Sundberg B. Cell-specific chemotyping and multivariate imaging by combined FT-IR microspectroscopy and orthogonal projections to latent structures (OPLS) analysis reveals the chemical landscape of secondary xylem. *Plant J* 2011; 66:903-14.
- [29] Mazurek S, Mucciolo A, Humbel BM, Nawrath C. Transmission fourier transform infrared microspectroscopy allows simultaneous assessment of cutin and cell-wall polysaccharides of *Arabidopsis* petals. *Plant J* 2013, 74, 880-91.
- [30] Yee N, Benning LG, Phoenix VR, Ferris FG. Characterization of metal-cyanobacteria sorption reactions: A combined macroscopic and infrared spectroscopic investigation *Environ Sci Technol* 2004;38, 775-82.
- [31] Shun-hui Yu, Hong-ping Deng, Bo Zhang, Zheng-xue Liu, Jun-jie Lin, Li Sheng, Jie Pan, Lianqi Huang & Jun-sheng Qi Physiological response of *Vetiveria zizanioides* to cadmium stress revealed by Fourier Transform Infrared Spectroscopy. *Spectroscopy Letters* 2017. DOI: 10.1080/00387010.2017.1355321
- [32] Hsiao TC, Xu LK, Sensitivity of growth of roots versus leaves to water stress: biophysical analysis and relation to water transport, *Journal of Experimental Botany* 2000;51(350): 1595-616.
- [33] Shao HB, Chu LY, Jaleel CA, Zhao CX. Water-deficit stress-induced anatomical changes in higher plants, *Comptes Rendus Biologies*, 2008; 331(3):215-25
- [34] Li XY, Liu DM, Wang J, Jian SG. Morphological, biochemical and physiological responses of a tropical coastal plant *Guettarda speciosa* to salt stress. *Global Ecology and Conservation* 2021; 32:e01887,